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Discordant Growth of Monozygotic Twins Starts at the Blastocyst Stage: A Case Study

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SUMMARY

Discordant growth is a common complication of monochorionic/diamniotic pregnancies; in approximately 50% of cases, the cause is unknown. The case presented here suggests that discordant growth of monozygotic twins could start during preimplantation development. Two inner cell masses (ICMs) within the same blastocyst may originate in uneven splitting of a single “parental” ICM, or the two ICMs may be formed independently *de novo*. We studied the transcriptomes of two morphologically distinct ICMs within a single blastocyst using high-resolution RNA sequencing. The data indicated that the two ICM were at different stages of development; one was in the earliest stages of lineage commitment, while the other had already differentiated into epiblast and primitive endoderm. IGF1-mediated signaling is likely to play a key role in ICM growth and to be the major driver behind these differences.

INTRODUCTION

Monozygotic (MZ) twins originate from the same conception event and are genetically identical. Approximately 80% of MZ twins originate from monochorionic/diamniotic pregnancies, meaning that they share the placenta and chorion, whereas each twin has its own amnion. These monochorionic/diamniotic MZ twins form if the inner cell mass (ICM), the part of the embryo that will give rise to the fetal body, splits at the early blastocyst stage, 4–6 days post-fertilization. Molecular events leading to this splitting are still in the domain of speculation. Time-lapse cinematography suggests that, at least *in vitro*, the reason might be purely mechanical; twins have been shown to form after blastocyst collapse and re-expansion, splitting the ICM into two groups of cells (Payne et al., 2007).

It is well documented that the incidence of MZ twins is higher following assisted reproduction techniques (Blickstein et al., 2003; Vitthala et al., 2009; Knopman et al., 2014; Tocino et al., 2015); however, we have rarely seen a good-quality blastocyst with two distinct ICMs among embryos donated for research in our center. High-resolution single-cell transcriptome analysis of the temporal and spatial patterns of gene expression in the human preimplantation embryos has been reported (Xue et al., 2013; Yan et al., 2013), and when we recently encountered such an embryo, we took the opportunity to compare the ICMs at the transcriptome level using next-generation sequencing (NGS).

RESULTS

The expanded day 6 blastocyst displayed two distinct ICMs 4 hr post-thawing, (Figure 1A). Single cells and cell borders were clearly visible in ICM1, whereas the cells in ICM2 were more tightly packed and no single cell could be distinguished, indicating that ICM2 is either better quality or more advanced than ICM1. To further explore this morphological discrepancy, we separated ICM1 and ICM2 from trophectoderm (TE) and by NGS evaluated the transcriptome of each of the three fractions. Tiered quality-control (QC) steps suggested good quality of cDNA and the library (Figure S1), as well as high homogeneity between samples (Figure S2). Nevertheless, we are aware that comparing two conditions with no replicates is not ideal and strong events and false positives may be missed.

Unexpectedly, the ICM1 and ICM2 transcriptome profiles did not cluster together; the ICM1 transcriptome was more similar to TE than to ICM2 (Figures 1B–1E). We found that ICM1 and ICM2 have 842 differently regulated genes with a fold change ≥ 2 and p value ≤ 0.5 , whereas the difference between ICM1 and TE was only in 128 genes and ICM2 and TE differed in expression of 356 genes. Pathway analyses drew our attention to insulin-like growth factor 1 (IGF1) signaling (Figures 2A and 2B). *IGF1* expression was relatively high in ICM2 and nearly undetectable in ICM1 (Figure 2C).

IGF1 acts through multiple pathways, including RAS/RAF/MAPK and PI3K/AKT. We therefore investigated whether the expression levels of downstream mediators,

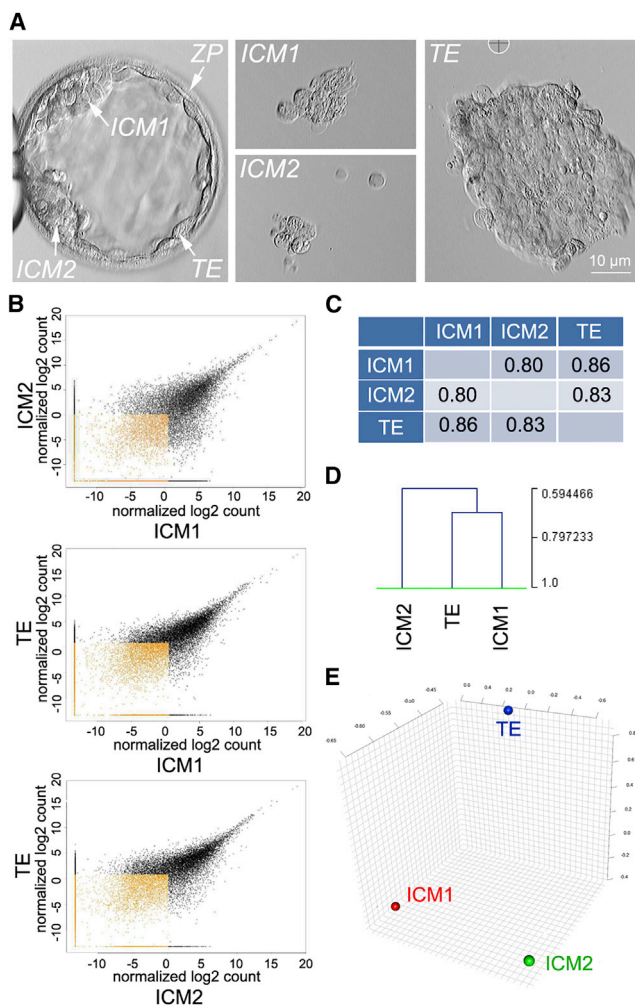


Figure 1. Transcriptome of Two ICMs of the MZ Twin Embryo Does Not Cluster Together

(A) Two ICMs (ICM1 and ICM2) of a day 6 natural MZ twin embryo mechanically separated from TE.

(B–E) Sample clustering used correlation of normalized read counts of expressed genes and average linkage clustering. Comparisons of normalized gene counts (B), correlation (C), clustering based on correlation (D), and principal component analysis (E) demonstrate that ICM1 is closer to TE than to ICM2.

members of RAF, PI3K, and AKT families, are different between ICM1 and ICM2 (Figures 2C and S3). Interestingly, from three RAF family members, *BRAF* was specifically expressed in ICM2, whereas *ARAF* was expressed only in TE. *RAF1* was found in all three fractions, at a level twice as high in ICM1 as in ICM2. Among class I PI3K catalytic subunits, only *PIK3CA* was expressed almost exclusively in ICM2. *PIK3CB* and *PIK3CD* were expressed in all three fractions, whereas *PIK3CG* was only in TE. Expression levels of AKT family members were also cell fraction specific: *AKT3* was predominantly expressed in ICM2, whereas

AKT1 was expressed in ICM1 and TE and was barely detectable in ICM2. *AKT2* was expressed in all three fractions but at a higher level in ICM1 than in ICM2. Although we do not know whether the protein expression parallels mRNA levels and, moreover, whether the signaling cascade is active, the data suggest that ICM1 is deficient for IGF1-mediated signaling. These data may indicate that the IGF1 pathway in the ICM of the human preimplantation embryos is linked to specific members of the RAF, PI3K, and AKT families.

The differences between ICM1 and ICM2 in global gene expression and particularly in expression of IGF1 pathway members suggested that they are distinct not only morphologically (Figure 1A) but also on a molecular level. Next, we examined relative expression levels of ICM and TE markers (Yan et al., 2013). ICM markers *GDF3*, *KLF4*, and *POU5F1* were higher in ICM2, whereas *NANOG* was higher in ICM1 (Figure 3A). TE markers showed a more interesting pattern (Figure 3B); although all were expressed in TE, only *CDX2* was TE specific. *CLDN10* and *GATA2* were expressed around the same level in ICM1 as in TE, whereas they were barely detectable in ICM2. *TET2* was expressed in all three fractions, the highest in TE and the lowest in ICM2. Expression of TE markers in ICM1 and not in ICM2 raises the possibility of cross-contamination of the ICM1 fraction with TE cells during the isolation process. Although we cannot exclude this, a contamination with TE cells is highly unlikely—*CDX2* expression in ICM1 would parallel a pattern of other TE markers, and it does not. In contrast to in the mouse, in the human *CDX2* is first detected in day 5 human embryos and is exclusively expressed in TE (Niakan and Egan, 2013). Therefore, together with the data from the IGF1 pathway, these results strongly indicate that ICM1 and ICM2 were at different stages of development.

The first cell fate decision occurs during the transition from morula to blastocyst. The allocation of cells to the ICM occurs through cell divisions in which outside cells divide in an orientation that directs one daughter inward (Johnson and Ziomek, 1981). The ICM has been shown not to be a homogenous group of cells (Chazaud et al., 2006), and it is to be expected that at the initial stages of segregation, some of the very early ICM cells retain detectable levels of expression of the proteins present in parental cells. In an early day 5 blastocyst with a small but morphologically distinct ICM, we were able to detect within the ICM the cells expressing *NANOG* and *GATA2* (Figure 3C). To explore further the possibility that ICM1 is at a stage of development earlier than that of ICM2, we investigated the expression pattern of *YAP1* (Figure 3D). *YAP1* can shuttle between the cytoplasm and the nucleus, and its nuclear localization is linked to an active role in the TAED4-mediated induction of *CDX2* expression (Lorthongpanich



A KEGG Pathway Analysis on Regulated Genes

Significant Pathways (6)

Pathway Description (KEGG)	Nb Genes in Pathway	Nb Regulated Genes (Up / Down)	P-Value (All)	P-Value (Up)	P-Value (Down)	Min P-Value
Glioma	83	9 (4/5)	4.16E-03	NA	2.47E-02	4.16E-03
N-Glycan biosynthesis	46	6 (0/6)	4.04E-02	4.25E-03	NA	4.25E-03
ErbB signaling pathway	87	10 (6/4)	9.49E-03	NA	NA	9.49E-03
Progesterone-mediated oocyte maturation	86	6 (0/6)	NA	NA	1.75E-02	1.75E-02
Prostate cancer	89	9 (4/5)	3.04E-02	NA	NA	3.04E-02
Amino sugar and nucleotide sugar metabolism	44	8 (3/3)	3.42E-02	NA	NA	3.42E-02

Pathway Description (KEGG)	FAST DB STABLE ID	Gene Symbol	Regulation	Fold-Change	P-Value
Glioma	GSKG0017810	CALM2	down	6	NA
	GSKG0000351	SOS2	up	inf	NA
	GSKG0028642	BRAF	down	93	NA
	GSKG0010191	IGF1R	up	inf	3.00E+00
	GSKG0013057	TP53	up	79	4.00E+00
	GSKG0025452	E2F3	down	inf	4.00E+00
N-Glycan biosynthesis	GSKG0007722	ICF1	down	inf	4.00E+00
	GSKG0021342	PIK3CA	down	77	5.00E+00
	GSKG0012740	PRKCA	up	471	NA
	GSKG0022279	ALG3	up	inf	3.00E+00
	GSKG0000584	ALG6	up	88	3.00E+00
	GSKG0005837	CANAB	up	105	3.00E+00
ErbB signaling pathway	GSKG0021775	RTT1	up	inf	NA
	GSKG0031755	ALG13	up	19	3.00E+00
	GSKG0010522	NAGPA-AS1	up	inf	2.00E+00
	GSKG0002768	ABL2	up	inf	2.00E+00
	GSKG0009351	SOS2	up	inf	NA
	GSKG0028642	BRAF	down	93	NA
Progesterone-mediated oocyte maturation	GSKG0016739	NOX2	up	914	NA
	GSKG0021342	PIK3CA	down	77	5.00E+00
	GSKG0013418	STAT5B	up	inf	3.00E+00
	GSKG0006740	ERBB3	up	214	4.00E+00
	GSKG0012740	PRKCA	up	471	NA
	GSKG0032009	ELK1	down	672	NA
Prostate cancer	GSKG0014803	PRK4	down	147	2.00E+00
	GSKG0028642	BRAF	down	93	NA
	GSKG0009633	HSP90AA1	down	4	NA
	GSKG0013509	CDC27	down	31	NA
	GSKG0032011	RPS6KA3	down	44	4.00E+00
	GSKG0007722	ICF1	down	inf	4.00E+00
Amino sugar and nucleotide sugar metabolism	GSKG0021342	PIK3CA	down	77	5.00E+00
	GSKG0009351	SOS2	up	inf	NA
	GSKG0028642	BRAF	down	93	NA
	GSKG0008441	LINC00598	up	inf	3.00E+00
	GSKG0010191	IGF1R	up	inf	3.00E+00
	GSKG0013057	TP53	up	79	4.00E+00

B REACTOME Pathway Analysis on Regulated Genes

Significant Pathways (4)

Pathway Description (REACTOME)	Nb Genes in Pathway	Nb Regulated Genes (Up / Down)	P-Value (All)	P-Value (Up)	P-Value (Down)	Min P-Value
Metabolism of lipids and lipoproteins	150	8 (0/8)	NA	2.67E-02	NA	2.67E-02
Hemostasis	235	15 (0/15)	3.31E-02	NA	NA	3.31E-02
Signaling by BMP	23	4 (2/2)	4.45E-02	NA	NA	4.45E-02
APC-Cdc20 mediated degradation of Nrk2A	23	3 (0/3)	NA	NA	4.57E-02	4.57E-02

Pathway Description (REACTOME)	FAST DB STABLE ID	Gene Symbol	Regulation	Fold-Change	P-Value
Metabolism of lipids and lipoproteins	GSKG0029241	SCLE	up	21	2.00E+00
	GSKG0024764	HMGCS1	up	106	NA
	GSKG0029948	DGAT1	up	564	4.00E+00
	GSKG0017455	SDC1	up	inf	5.00E+00
	GSKG0027335	PPAP2B	up	286	3.00E+00
	GSKG0016924	GPI2	up	17	4.00E+00
Hemostasis	GSKG0020742	ABHD5	up	236	4.00E+00
	GSKG0016311	HADHB	up	678	NA
	GSKG0017610	CALM2	down	6	NA
	GSKG0004845	SERPINC1	down	1820	2.00E+00
	GSKG0026802	BRP3	up	inf	2.00E+00
	GSKG0017288	INPP5D	up	inf	4.00E+00
Signaling by BMP	GSKG0011134	ITGAM	down	inf	NA
	GSKG0021913	CD47	up	inf	NA
	GSKG0021342	PIK3CA	down	77	5.00E+00
	GSKG0007722	ICF1	down	inf	4.00E+00
	GSKG002367	SLC16A1	up	55	NA
	GSKG0007528	ITGAS	up	inf	NA
APC-Cdc20 mediated degradation of Nrk2A	GSKG0031987	PIR	up	358	NA
	GSKG0012740	PRKCA	up	471	NA
	GSKG0024046	F2R	down	inf	3.00E+00
	GSKG0000242	SLC7A8	up	24	NA
	GSKG0004864	PLCB3	up	inf	4.00E+00
	GSKG0018555	BMP2	down	18	NA

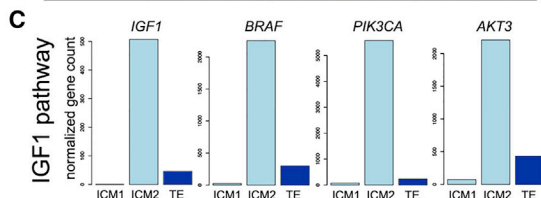


Figure 2. Pathway Analysis of Genes Differently Expressed between ICM1 and ICM2 Highlighted the IGF1 Signaling Pathway

(A) Kyoto Encyclopedia of Genes and Genomes database pathway analysis on regulated genes highlighted six pathways. Three of the six pathways involve the IGF1/BRAF/PIK3CA signaling cascade. (B) Reactome database pathway analysis on regulated genes highlighted four pathways, from which one involves the IGF1/PIK3CA signaling cascade. (C) Expression of *IGF1* and downstream mediators *BRAF*, *AKT3*, and *PIK3CA* of its signaling pathway is higher in ICM2 than in ICM1 or TE.

et al., 2013; Home et al., 2012; Nishioka et al., 2009). We found that ICM1 was expressing higher levels of *YAP1* than those of not only ICM2 but also TE, which was somewhat discordant with the published mouse data. Therefore, we investigated the *YAP1* expression pattern in morula to late blastocyst stages of human embryo development. In contrast to the mouse, we found that *YAP1* is heavily expressed and localized in the nuclei of ICM cells in the early stage blastocyst, whereas at later stages, similarly to the mouse, the expression in the ICM is exclusively cytoplasmic and seems to be dwindling in both the ICM and most TE cells. Our immunostaining data support the hypothesis that ICM1 and ICM2 are at different stages of the development, with ICM2 being developmentally more advanced than ICM1.

To test this further, we looked into the expression of epiblast (Epi) and primitive endoderm (PE) markers (Yan et al., 2013), which would be expressed only in an advanced ICM. The Epi markers *FOXD3*, *PRDM14*, and *UTF1* were found almost exclusively in ICM2. All PE markers were found in both ICMs, although their expression was higher in ICM2, especially *DPPA5* and *IFITM1* (Figure 3E).

If *BRAF*, *PIK3CA*, and *AKT3* are specific for IGF1 downstream signaling at that stage of development, as the data from the ICM2 sample suggested (Figure 4A), their expression would show a similar pattern in other embryos. We pooled together the same type of samples: ICM pool (ICMP; n = 3) and TE pool (TEP; n = 3) from three late blastocysts and analyzed them using a different RNA sequencing (RNA-seq) platform (the SOLiD platform from Life Technologies instead of Illumina). We understand that the data obtained from two different platforms are not directly comparable. However, the data obtained should reflect the biology of the samples, and if the *BRAF/PIK3CA/AKT3* signaling pathway or pathways are specific for ICM, their expression should show a similar trend regardless of the RNA-seq platform used. The data from the SOLiD platform suggested that may not be the case (Figures 4B–4F and S4). None of *RAF* and *AKT* family members showed strong ICM specificity as detected in

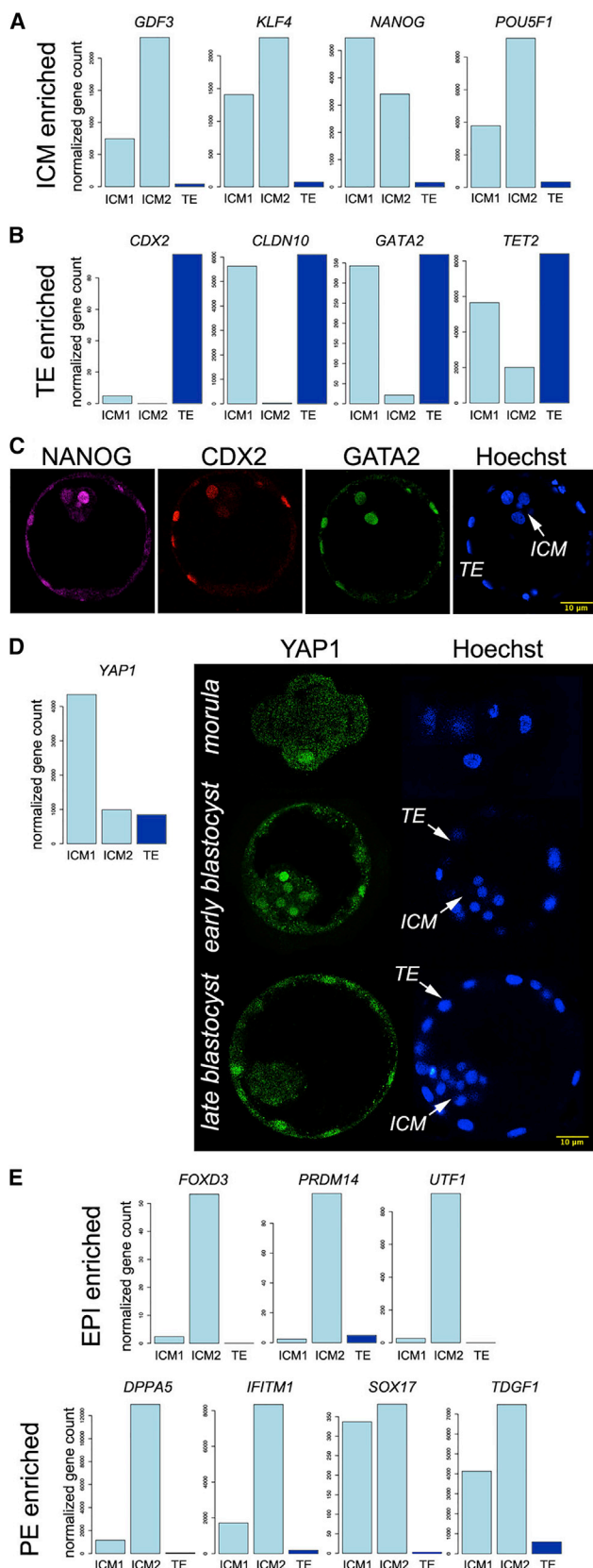


Figure 3. Lineage Marker Expression Pattern Suggests that ICM2 Is Developmentally More Advanced Than ICM1

(A) Genes known to have significantly higher expression in ICM (*GDF3*, *KLF4*, *NANOG*, and *POU5F1*) show the expected pattern.

(B) Among four genes known to have significantly higher expression in TE (*CDX2*, *CLDN10*, *GATA2*, and *TET2*), two of them, *CLDN10* and *GATA2*, are equally high expressed in ICM1 and in TE.

(C) At the early blastocyst stage, some cells still have dual expression of ICM and TE markers. A confocal image shows a day 5 blastocyst immunostained for the ICM marker *NANOG* and TE markers *CDX2* and *GATA2*.

(D) Expression of transcription factor *YAP1*, which plays a role in initiating *CDX2* expression and committing cells to the TE lineage, is higher in ICM1 than in ICM2. *YAP1* immunostaining of preimplantation embryos of different stages of development shows that *YAP1* is detected in the nuclei of ICM cells only at the early stage of blastocyst formation. At the late blastocyst stage, *YAP1* nuclear localization is restricted to TE.

(E) Expression of Epi and PE markers is higher in ICM2 than in ICM1, suggesting that ICM2 is developmentally more advanced than ICM1.

ICM2. In the newly analyzed samples, *PIK3CD* and *PIK3CG* seemed to be more specific to ICM than was *PIK3CA*, which stood out from ICM2 analyses.

DISCUSSION

MZ twins are recognized to be at greater risk of congenital anomalies and other morbidity than dizygotic ones (Lewi et al., 2008). One of the most common complications is severe discordant growth, which is defined as 25% or greater difference in birth weight for live-born twins or cases resulting in intrauterine fetal death and 25% or greater difference in estimated fetal weight in the absence of twin-to-twin transfusion syndrome. Such severe discordant growth has been reported in 7%–14% of all monochorionic/diamniotic pregnancies (Harper et al., 2013; Lewi et al., 2008; Acosta-Rojas et al., 2007; Fick et al., 2006). Hidden loss of such pregnancies is not known. Unequal placental sharing, defined as a placental territory discordance of ≥ 1.5 or as one twin receiving blood from $>60\%$ of the placenta, can explain about 50% of pregnancies with discordant growth; the cause of the other half in most cases is unknown (Lewi et al., 2007; Fick et al., 2006).

NGS and immunostaining data in this study suggest that discordant growth of MZ twins might start as early as the blastocyst stage (Figure 4A). Tocino et al. (2015) reported a case of asymmetric MZ twinning after a single embryo transfer. Discordant growth was observed by ultrasound as early as 24 days following the embryo transfer, and a difference in size was constant throughout the pregnancy. The prevalence of this early stage discordant growth is



A

Markers		ICM1	ICM2	TE
ICM-enriched	<i>GDF3</i>	+	+++	–
	<i>KLF4</i>	++	+++	–
	<i>NANOG</i>	+++	++	–
	<i>POU5F1</i>	+	+++	–
TE-enriched	<i>CDX2</i>	–	–	+++
	<i>CLDN10</i>	+++	–	++
	<i>GATA2</i>	+++	–	+++
	<i>TET2</i>	++	+	+++
	<i>YAP1</i>	+++	+	+
EPI-enriched	<i>FOXD3</i>	–	+++	–
	<i>PRDM14</i>	–	+++	–
	<i>UTF1</i>	–	+++	–
PE-enriched	<i>DPPA5</i>	+	+++	–
	<i>IFITM1</i>	+	+++	–
	<i>SOX17</i>	+++	+++	–
	<i>TDGF1</i>	++	+++	–
IGF1-mediated signaling	<i>IGF1</i>	–	+++	+
	<i>RAF</i>	<i>ARAF</i>	–	+++
		<i>BRAF</i>	–	+++
		<i>RAF1</i>	+++	++
	<i>PIK3C</i>	<i>PIK3CA</i>	–	+++
		<i>PIK3CB</i>	+	+++
		<i>PIK3CD</i>	+	+++
		<i>PIK3CG</i>	–	+++
	<i>AKT</i>	<i>AKT1</i>	+++	–
		<i>AKT2</i>	+++	+
		<i>AKT3</i>	–	+++

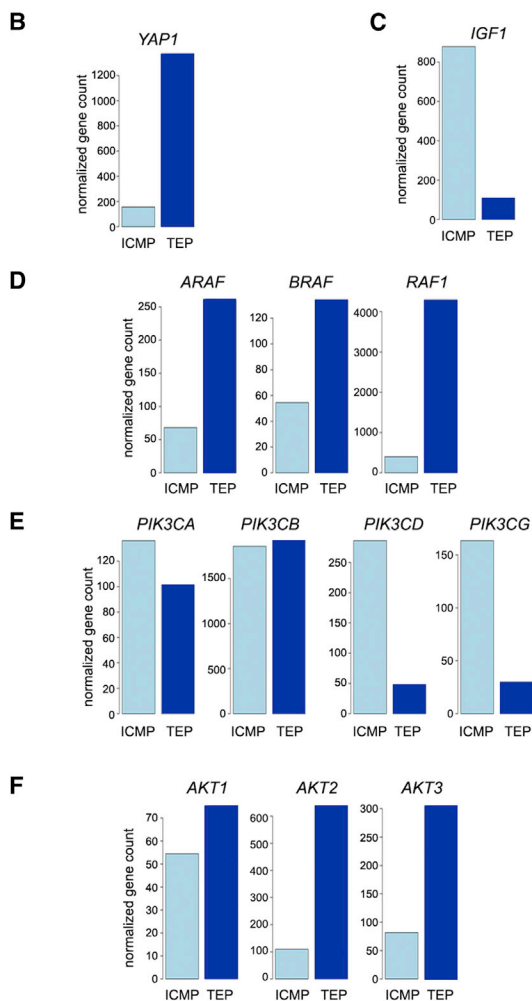


Figure 4. BRAF/PIK3CA/AKT3 Signaling Is Not Specific for the ICM of Late Blastocysts

(A) Gene expression pattern of lineage markers and IGF1-mediated signaling pathway is shown in three fractions of the MZ twin blastocyst, ICM1, ICM2, and TE.

(B) High *YAP1* expression levels in TEP ($n = 3$) and low levels in ICMP ($n = 3$) confirm that the samples used for these analyses are isolated from the late stage blastocysts.

(C–F) Although IGF1 expression is high in ICMP (C), downstream mediators BRAF (D), PIK3CA (E), and AKT3 (F), exclusively expressed in ICM2 (A), do not show an ICM-specific pattern in ICMP ($n = 3$).

not known, and further case reports and studies are needed. Interestingly, the couple who donated the embryo for this study had been through multiple unsuccessful in vitro fertilization cycles, and an inherited pathology as causative of the discordant development observed remains as a distant possibility.

Our studies suggest that the role of IGF1-mediated signaling in ICM growth is worthy of further exploration and may shed light on molecular pathways controlling early human embryo development. IGF1, also known as somatomedin C, induces growth and metabolism of preimplantation embryos in vitro (Harvey and Kaye, 1992). Studies in multiple species have demonstrated that IGF1 selectively stimulates the proliferation of cells in an ICM, as well as protecting them from apoptosis (Green and Day, 2013; Thieme et al., 2012; Bonilla et al., 2011; Velazquez et al., 2009; Kimber et al., 2008; Spanos et al., 2000; Lighten et al., 1998). Recent reports have correlated levels of IGF1 in follicular fluid with both quality of embryos and clinical pregnancy during in vitro fertilization cycles (Mehta et al., 2013). However, preimplantation embryos exposed to high non-physiological concentrations of IGF1 undergo extensive apoptosis of the ICM nuclei (Chi et al., 2000; Makarevich and Markkula, 2002; Velazquez et al., 2011), suggesting that IGF1 signaling in preimplantation embryos is tightly regulated. In light of these reports, including our findings, the possibility of adding IGF1 into embryo culture media during assisted reproduction to improve the odds for better-quality embryos and fetal growth post-transfer should be explored further, and downstream pathways and molecular mechanisms should be investigated.

The different stages of development of the two ICMs might result from an uneven split of an initial single ICM during blastocyst collapse. A delay in formation of intercellular adhesions within the ICM might make the ICM prone to splitting (Togashi et al., 2015). The smaller part, in this case ICM1, might be lagging behind due to a lack of critical cell mass. However, we cannot exclude the possibility that ICM1 and ICM2 formed independently de novo at



different time points (Noli et al., 2015). TE cells from the human non-expanded blastocysts are not fully committed; isolated and reaggregated TE cells are able to develop into blastocysts with ICM cells expressing NANOG (De Paepe et al., 2013). Either way, our case report suggests new avenues to studies of cellular and molecular events behind MZ twinning and discordant growth, commonly associated with monochorionic/diamniotic pregnancies, as well as lineage commitment at the earliest stages of human development.

EXPERIMENTAL PROCEDURES

The work described here was done under license from the UK Human Fertilization and Embryology Authority (research license R0075) and has local ethical approval (UK National Health Service Research Ethics Committee, reference 06/Q0702/90). Informed consent was obtained from all subjects, and the experiments conformed to the principles set out in the WMA Declaration of Helsinki and the NIH Belmont Report. No financial inducements were offered for donation.

Human Embryo Culture

The embryos were thawed using Quinn's Advantage thaw kit according to the manufacturer's instructions (Sage). After thawing, the embryos were cultured in 40- μ l microdrops of Quinn's blastocyst medium supplemented with 10% synthetic protein serum substitute (Sage) under mineral oil (Sage) at 37°C in 5% CO₂ and 5% O₂ in a humidified atmosphere. ICM and TE were separated mechanically as described (Capalbo et al., 2013).

RNA-Seq and Bioinformatics

TE and ICM fractions of the MZ twin embryo were lysed in 1 μ l of lysis buffer A from the Prelude direct lysis module (NuGEN), and cDNA was synthesized and amplified using the Ovation RNA-seq system V2 enabled by proprietary ribo-single primer isothermal amplification technology following the manufacturer's instructions (NuGEN). The cDNA was purified using the MinElute reaction cleanup kit (QIAGEN), and it was analyzed using the 2100 Bioanalyzer with the DNA 1000 LabChip and the 2200 TapeStation (Agilent). Unidirectional strand-specific 450–500 bp libraries were constructed on the Apollo 324 NGS system (IntergenX) and then sequenced on the HiSeq 2000 system (Illumina) by Oxford Gene Technology. The libraries were quantified using Qubit (Life Technologies), and the size profile was analyzed on the 2200 TapeStation (Agilent). Reads were aligned to the human genome (hg19) using Spliced Transcripts Alignment to a Reference v.2.4.0 (Dobin et al., 2013).

The cDNA was prepared from TE and ICM fractions pooled from three normal embryos using the same protocol. The library construction and the NGS on SOLiD platform (Life Technologies) were performed by Microsynth, and the alignments to hg19 were performed using LifeScope v.2.5 software (<http://www.thermofisher.com/uk/en/home/technical-resources/software-downloads/lifescopegenomic-analysis-software.html>).

GenoSplice Technology performed QC, processing, and further analyses of the data. For each gene present in the Friendly Alterna-

tive Splicing and Transcripts Database v.2014_2, reads aligning on constitutive regions (that are not prone to alternative splicing) were counted. Based on these read counts, normalization and differential gene expression were performed using DESeq (v.1.16.0 on R v.3.1.3) (Anders and Huber, 2010). Sample clustering was performed with MeV 4.9 (Saeed et al., 2003) using correlation of normalized read counts of expressed genes and average linkage clustering.

Immunostaining

Immunostaining was performed as described (Niakan and Egan, 2013). Primary antibodies were rabbit polyclonal anti-YAP1 (Cell Signaling Technology, Catalog No. 4912) and anti-GATA2 (Santa Cruz Biotechnology, catalog no. SC-9008), goat polyclonal anti-NANOG (R&D Systems, catalog no. AF1997), and mouse monoclonal anti-CDX2 (BioGenex, catalog no. MU392A-UC). Secondary antibodies were purchased from Life Technologies. Nuclei are visualized with DAPI (Vector Laboratories). Confocal microscopy was carrying out using a Leica TCS SP5 microscope. Images were process using ImageJ64 and Adobe Photoshop Creative Cloud 2014 software.

ACCESSION NUMBERS

The accession number for the RNA-seq data reported in this paper is Sequence Read Archive: SRP063754.

SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures and can be found with this article online at <http://dx.doi.org/10.1016/j.stemcr.2015.10.006>.

AUTHOR CONTRIBUTIONS

L.N. and A.C. performed experiments. L.N., C.O., Y.K., and D.I. contributed to the design of the study and took part in the analysis of the results and writing the manuscript.

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